

Piccolo® General Chemistry 6



For In Vitro Diagnostic Use and Healthcare
Professional Use Only
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Applicable to US customers only
CLIA Waived: Use lithium heparin whole blood, only
Moderate Complexity: Use lithium heparin whole blood, lithium heparin plasma, or serum



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1. Intended Use

The Piccolo® General Chemistry 6, used with the Piccolo Xpress® chemistry analyzer, is intended to be used for the *in vitro* quantitative determination of alanine aminotransferase (ALT), aspartate aminotransferase (AST), creatinine, gamma glutamyltransferase (GGT), glucose, and blood urea nitrogen (BUN) in heparinized whole blood, heparinized plasma, or serum in a clinical laboratory setting or point-of-care location/ near patient testing.

For US Customers Only

The tests on this panel are waived under CLIA '88 regulations. If a laboratory modifies the test system instructions, then the tests are considered high complexity and subject to all CLIA requirements. For CLIA waived labs, only lithium heparin whole blood may be tested. For use in moderate complexity labs, lithium heparinized whole blood, lithium heparinized plasma, or serum may be used.

A CLIA Certificate of Waiver is needed to perform CLIA waived testing. A Certificate of Waiver can be obtained from the Centers for Medicare & Medicaid Services (CMS).

2. Summary and Explanation of Tests

The Piccolo General Chemistry 6 reagent disc and the Piccolo Xpress chemistry analyzer comprise an *in vitro* diagnostic system that aids the physician in diagnosing the following disorders in adults:

Alanine Aminotransferase (ALT):	Liver diseases, for example, viral hepatitis and cirrhosis.
Aspartate aminotransferase (AST):	Liver disease, for example, hepatitis and viral jaundice, shock liver.
Creatinine:	Renal disease and renal dialysis.
Gamma glutamyltransferase (GGT):	Liver diseases, for example, alcoholic cirrhosis and primary and secondary liver tumors.
Glucose:	Carbohydrate metabolism disorders, for example, adult and juvenile diabetes mellitus and hypoglycemia.
Blood urea nitrogen (BUN):	Renal diseases.

As with any diagnostic test procedure, all other test procedures including the clinical status of the patient, should be considered prior to final diagnosis.

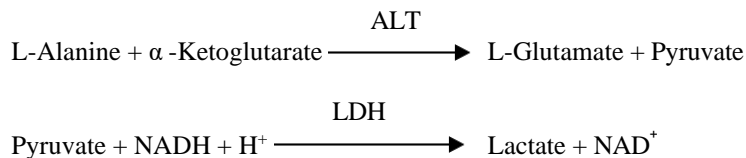
3. Principles of Method

Alanine Aminotransferase (ALT)

Alanine aminotransferase (ALT) has been measured by three methods. Two of these methods—the colorimetric dinitrophenylhydrazine coupling technique^{1,2} and the fluorescent enzymatic assay—are rarely used.³ An enzymatic method based on the work of Wróblewski and LaDue⁴ is the most common technique for determining ALT concentrations in serum.

A modified Wróblewski and LaDue procedure has been proposed as the recommended procedure of the International Federation of Clinical Chemistry (IFCC).⁵

The method developed for use on the Piccolo analyzers is a modification of the IFCC-recommended procedure. In this reaction, ALT catalyzes the transfer of an amino group from L-alanine to α -ketoglutarate to form L-glutamate and pyruvate. Lactate dehydrogenase catalyzes the conversion of pyruvate to lactate. Concomitantly, NADH is oxidized to NAD^+ , as illustrated in the following reaction scheme.

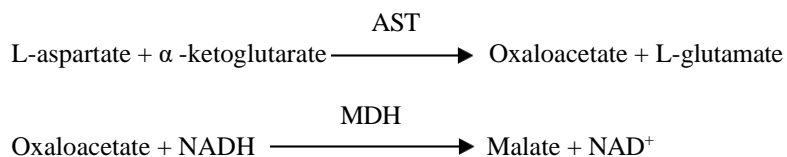


The rate of change of the absorbance difference between 340 nm and 405 nm is due to the conversion of NADH to NAD^+ and is directly proportional to the amount of ALT present in the sample.

Aspartate Aminotransferase (AST)

The aspartate aminotransferase (AST) test is based on the Karmen rate method⁶ as modified by Bergmeyer.⁷ The current International Federation of Clinical Chemistry (IFCC) reference method utilizes the Karmen/Bergmeyer technique of coupling malate dehydrogenase (MDH) and reduced nicotinamide dinucleotide (NADH) in the detection of AST in serum.^{7,8} Lactate dehydrogenase (LDH) is added to the reaction to decrease interference caused by endogenous pyruvate.

AST catalyzes the reaction of L-aspartate and α -ketoglutarate into oxaloacetate and L-glutamate. Oxaloacetate is converted to malate and NADH is oxidized to NAD^+ by the catalyst MDH.

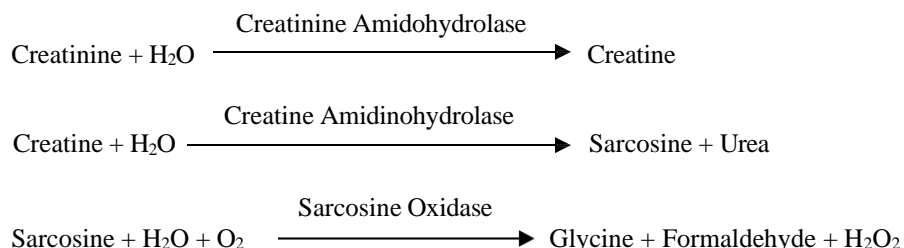


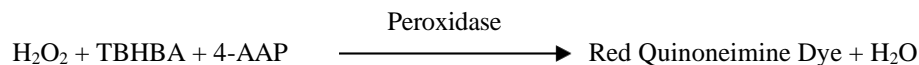
The rate of absorbance change at 340 nm/405 nm caused by the conversion of NADH to NAD^+ is directly proportional to the amount of AST present in the sample.

Creatinine (CRE)

The Jaffe method, first introduced in 1886, is still a commonly used method of determining creatinine levels in blood. The current reference method combines the use of fuller's earth (floridin) with the Jaffe technique to increase the specificity of the reaction.^{9,10} Enzymatic methods have been developed that are more specific for creatinine than the various modifications of the Jaffe technique.^{11,12,13} Methods using the enzyme creatinine amidohydrolase eliminate the problem of ammonium ion interference found in techniques using creatinine iminohydrolase.¹⁴

In the coupled enzyme reactions, creatinine amidohydrolase hydrolyzes creatinine to creatine. A second enzyme, creatine amidinohydrolase, catalyzes the formation of sarcosine from creatine. Sarcosine oxidase causes the oxidation of sarcosine to glycine, formaldehyde and hydrogen peroxide (H_2O_2). In a Trinder finish, peroxidase catalyzes the reaction among the hydrogen peroxide, 2,4,6-tribromo-3-hydroxybenzoic acid (TBHBA) and 4-aminoantipyrine (4-AAAP) into a red quinoneimine dye. Potassium ferrocyanide and ascorbate oxidase are added to the reaction mixture to minimize the potential interference of bilirubin and ascorbic acid, respectively.





Two cuvettes are used to determine the concentration of creatinine in the sample. Endogenous creatine is measured in the blank cuvette, which is subtracted from the combined endogenous creatine and the creatine formed from the enzyme reactions in the test cuvette. Once the endogenous creatine is eliminated from the calculations, the concentration of creatinine is proportional to the intensity of the red color produced. The endpoint reaction is measured as the difference in absorbance between 550 nm and 600 nm.

eGFR (calculated)

Serum creatinine is routinely measured as an indicator of renal function. Because creatinine is influenced by age, gender and race, chronic kidney disease (CKD) may not be detected using serum creatinine alone. Thus, the National Kidney Disease Education Program strongly recommends that laboratories routinely report an estimated Glomerular Filtration Rate (eGFR) when serum creatinine is measured for patients 18 and older. Routinely reporting the eGFR with all serum creatinine determinations allows laboratories to help identify individuals with reduced kidney function and help facilitate the detection of CKD. Calculated eGFR values of <60 ml/min are generally associated with increased risk of adverse outcomes of CKD.

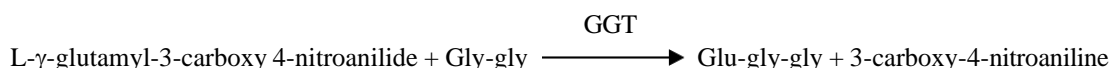
Calculation of the eGFR is performed by the Piccolo using the patient's age, gender and race. The Piccolo method for creatinine is traceable to the IDMS reference method for creatinine so that the following form of the MDRD equation for calculating the eGFR can be used.

$$\text{GFR (mL/min/1.73 m}^2\text{)} = 175 \times (\text{S}_{\text{cr}})^{-1.154} \times (\text{Age})^{-0.203} \times (0.742 \text{ if female}) \times (1.212 \text{ if African American})$$

Gamma Glutamyltransferase (GGT)

The first quantitative methods developed to measure gamma glutamyltransferase (GGT) involved a second reaction to form an azo dye that combined with a chromophore.^{15,16} The change to L-γ-glutamyl-*p*-nitroanilide as the substrate in the reaction eliminated the dye-formation step.¹⁷ Due to the poor solubility and stability of L-γ-glutamyl-*p*-nitroanilide, this procedure was modified to use the substrate L-γ-glutamyl-3-carboxy-4-nitroanilide.¹⁸ The International Federation of Clinical Chemistry (IFCC) recommended GGT method is based on the latter substrate, with glycylglycine as the other substrate.¹⁹

Abaxis has modified the IFCC method to react at 37°C. The addition of sample containing gamma glutamyl transferase to the substrates L-γ-glutamyl-3-carboxy-4-nitroanilide and glycylglycine (gly-gly) causes the formation of L-γ-glutamyl-glycylglycine (glu-gly-gly) and 3-carboxy-4-nitroaniline.

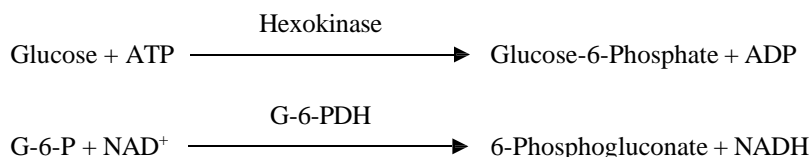


The absorbance of this rate reaction is measured at 405 nm. The production of 3-carboxy-4-nitroaniline is directly proportional to the GGT activity in the sample.

Glucose (GLU)

Measurements of glucose concentration were first performed using copper-reduction methods (such as Folin-Wu²⁰ and Somogyi-Nelson^{21,22}). The lack of specificity in copper-reduction techniques led to the development of quantitative procedures using the enzymes hexokinase and glucose oxidase. The glucose test incorporated into the Piccolo General Chemistry 6 reagent disc is a modified version of the hexokinase method, which has been proposed as the basis of the glucose reference method.²³

The reaction of glucose with adenosine triphosphate (ATP), catalyzed by hexokinase (HK), produces glucose-6-phosphate (G-6-P) and adenosine diphosphate (ADP). Glucose-6-phosphate dehydrogenase (G-6-PDH) catalyzes the reaction of G-6-P into 6-phosphogluconate and the reduction of nicotinamide adenine dinucleotide (NAD⁺) to NADH.

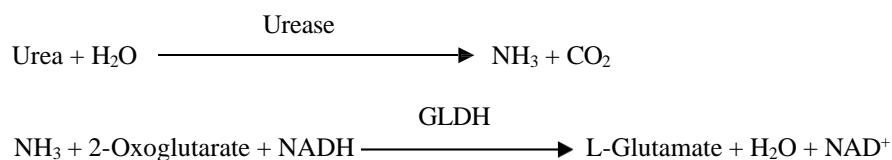


The absorbance is measured bichromatically at 340 nm and 850 nm. The production of NADH is directly proportional to the amount of glucose present in the sample.

Blood Urea Nitrogen (BUN)

Urea can be measured both directly and indirectly. The diacetyl monoxime reaction, the only direct method to measure urea, is commonly used but employs dangerous reagents.²⁴ Indirect methods measure ammonia created from the urea; the use of the enzyme urease has increased the specificity of these tests.²⁵ The ammonia is quantitated by a variety of methods, including nesslerization (acid titration), the Berthelot technique^{26,27} and coupled enzymatic reactions.^{28,29} Catalyzed Berthelot procedures, however, are erratic when measuring ammonia.³⁰ Coupled-enzyme reactions are rapid, have a high specificity for ammonia, and are commonly used. One such reaction has been proposed as a candidate reference method.³¹

In the coupled-enzyme reaction, urease hydrolyzes urea into ammonia and carbon dioxide. Upon combining ammonia with 2-oxoglutarate and reduced nicotinamide adenine dinucleotide (NADH), the enzyme glutamate dehydrogenase (GLDH) oxidizes NADH to NAD⁺.



The rate of change of the absorbance difference between 340 nm and 405 nm is caused by the conversion of NADH to NAD⁺ and is directly proportional to the amount of urea present in the sample.

4. Principles of Operation

Refer to the Piccolo Xpress chemistry analyzer Operator's Manual, for the Principles and Limitations of the Procedure.

5. Description of Reagents

Reagents

Each Piccolo General Chemistry 6 reagent disc contains dry test-specific reagent beads (described below). A dry sample blank reagent (comprised of buffer, surfactants, excipients, and preservatives) is included in each disc for use in calculating concentrations of alanine aminotransferase (ALT), aspartate aminotransferase (AST), gamma glutamyltransferase (GGT), glucose (GLU), and blood urea nitrogen (BUN). A dedicated sample blank is included in the disc for creatinine (CRE). Each reagent disc also contains a diluent consisting of surfactants, excipients, and preservatives.

Table 1: Reagents

Component	Quantity/Disc
Adenosine-5'-diphosphate,	4 µg
L-Alanine	874 µg
4-Aminoantipyrine HCl	14 µg
Ascorbate oxidase	0.3 U
L-Aspartic acid	426 µg
Creatine amidinohydrolase	3 U
Creatinine amidohydrolase	1 U
Glucose-6-phosphate dehydrogenase	0.05 U
L-Glutamic acid dehydrogenase	0.01 U
L-Glutamic acid γ-(3-carboxy-4-nitroanilide), ammonium salt	30 µg
Glycylglycine	317 µg
Hexokinase	0.1 U
α-Ketoglutarate, disodium salt	60 µg
α-Ketoglutaric acid	54 µg
Lactate dehydrogenase	0.002 U
Lactate dehydrogenase	0.1 U
Malate dehydrogenase (MDH)	0.01 U

Table 1: Reagents (continued)

Component	Quantity/Disc
β-Nicotinamide adenine dinucleotide, reduced (NADH)	18 µg
Peroxidase	0.7 U
Potassium ferrocyanide	0.9 µg
Sarcosine oxidase	0.7 U
2,4,6-Tribromo-3-hydroxybenzoic acid	188 µg
Urease	0.05 U
Adenosine-5'-diphosphate, monopotassium salt	4 µg
Adenosine-5'-triphosphate, disodium salt	12 µg
4-Aminoantipyrine	5.4 µg
3,5-Dichloro-2-hydroxybenzenesulfonic acid, Sodium salt (DHBSA)	34.8 µg
Ethylenediaminetetraacetic acid (EDTA), disodium salt	36 µg
Ethylenediaminetetraacetic acid (EDTA), Tetrasodium salt	201 µg
Alpha-Ketoglutaric acid, disodium salt	60 µg
Uricase	0 U
Magnesium acetate, tetrahydrate	5 µg

Warnings and Precautions

- For *In vitro* Diagnostic Use
- The diluent container in the reagent disc is automatically opened when the analyzer drawer closes. A disc with an opened diluent container cannot be re-used. Ensure that the sample or control has been placed into the disc before closing the drawer.
- Used reagent discs contain human body fluids. Follow good laboratory safety practices when handling and disposing of used discs.³² See the Piccolo Xpress chemistry analyzer Operator's Manual for instructions on cleaning biohazardous spills.
- The reagent discs are plastic and may crack or chip if dropped. **Never** use a dropped disc as it may spray biohazardous material throughout the interior of the analyzer.
- Reagent beads may contain acids or caustic substances. The operator does not come into contact with the reagent beads when following the recommended procedures. In the event that the beads are handled (e.g., cleaning up after dropping and cracking a reagent disc), avoid ingestion, skin contact, or inhalation of the reagent beads.
- This mixture does not contain substances assessed to be vPvB / PBT according to Regulation (EC) No 1907/2006, Annex XIII. This product contains components considered to have endocrine disrupting properties for environment, according to REACH Article 57(f), Regulation (EU) 2018/605 or Regulation (EU) 2017/2100. Refer to the Safety Data Sheet (SDS) for specific details

Instructions for Reagent Handling

Reagent discs may be used directly from the refrigerator without warming. Do not allow discs to remain at room temperature at 20-25°C (68-77°F) longer than 48 hours prior to use. Open the sealed foil pouch and remove the disc, being careful not to touch the bar code ring located on the top of the disc. Use according to the instructions provided in the Piccolo Xpress chemistry analyzer Operator's Manual. A disc not used within 20 minutes of opening the pouch should be discarded.

Storage

Store reagent discs in their sealed pouches at 2-8 °C (36-46 °F). Do not expose opened or unopened discs to direct sunlight or temperatures above 32 °C (90 °F). Reagent discs may be used until the expiration date included on the package. The expiration date is also encoded in the bar code printed on the bar code ring. An error message will appear on the Piccolo Xpress chemistry analyzer display if the reagents have expired.

Indications of Reagent Disc Instability/Deterioration

A torn or otherwise damaged pouch may allow moisture to reach the unused rotor and adversely affect reagent performance. Do not use a rotor from a damaged pouch.

6. Instrument

See Piccolo Xpress chemistry analyzer Operator's Manual for complete information on use of the analyzer.

7. Sample Collection and Preparation

Sample collection techniques are described in the "Sample Collection" section of the Piccolo Xpress chemistry analyzer Operator's Manual.

- The minimum required sample size is ~100 µL of heparinized whole blood, heparinized plasma, serum or control material. The reagent disc sample chamber can contain up to 120 µL of sample.
- Whole blood samples obtained by venipuncture must be homogeneous before transferring a sample to the reagent disc. Gently invert the collection tube several times just prior to sample transfer. Do not shake the collection tube; shaking may cause hemolysis.
- Whole blood samples should only be obtained via venipuncture, not from capillary blood.
- Whole blood venipuncture samples should be run within 60 minutes of collection.^{33,43} **Glucose** concentrations are affected by the length of time since the patient has eaten and by the type of sample collected from the patient. To accurately determine glucose results, samples should be obtained from a patient who has been fasting for at least 12 hours. The glucose concentration decreases approximately 0.28-0.67 mmol/L in 1 hour in uncentrifuged samples stored at room temperature.³⁴
- Refrigerating whole blood samples can cause significant changes in concentrations of **aspartate aminotransferase, creatinine** and **glucose**.³⁵ The sample may be separated into plasma or serum and stored in capped sample tubes at 2-8 °C (36-46 °F) if the sample cannot be run within 60 minutes.
- Use only lithium heparin evacuated specimen collection tubes for whole blood or plasma samples. Use no-additive evacuated specimen collection tubes or serum separator tubes for serum samples.
- Start the test within 10 minutes of transferring the sample into the reagent disc.

8. Procedure

Materials Provided

- One Piccolo General Chemistry 6 PN: 400-1006 (a box of discs PN: 400-0006)

Materials Required but not Provided

- Piccolo Xpress chemistry analyzer
- Sample transfer pipettes (fixed volume approximately 100 µL) and tips are provided with each Piccolo Xpress chemistry analyzer and may be reordered from Abaxis.
- Commercially available control reagents recommended by Abaxis (contact Abaxis Technical Service for approved control materials and expected values).
- Timer

Test Parameters

The Piccolo Xpress chemistry analyzer operates at ambient temperatures between 15 °C and 32 °C (59-90 °F). The analysis time for each Piccolo General Chemistry 6 reagent disc is less than 14 minutes. The analyzer maintains the reagent disc at a temperature of 37 °C (98.6 °F) over the measurement interval.

Test Procedure

The complete sample collection and step-by-step operating procedures are detailed in the Piccolo Xpress chemistry analyzer Operator’s Manual.

Calibration

The Piccolo Xpress chemistry analyzer is calibrated by the manufacturer before shipment. The bar code printed on the bar code ring provides the analyzer with disc-specific calibration data. See Piccolo Xpress chemistry analyzer Operator’s Manual.

The Piccolo Xpress chemistry analyzer is calibrated using internal calibrators or reference materials. The methods and materials used in the procedures for the calibration and control value assignments are traceable either to the comparative method and/or to the standards listed below:

- IFCC – International Federation of Clinical Chemistry
- NIST – National Institute of Standards and Technology
- CRMLN – Cholesterol Reference Method Laboratory Network
- SRM – Standard Reference Material
- CRM – Certified Reference Material

The calibration and value assignment processes are in compliance with EN ISO 17511, Metrological Traceability of Values Assigned to Calibrators and Control Materials.

The measured analytes in the General Chemistry 6 are traceable to the following reference materials or methods. The Piccolo Xpress Analysis System controls and calibration verification materials are validated for use only with the Piccolo Xpress Analysis System and assigned values may not be commutable with other methods. For specific analyte reference standard methods, refer to Section 3. Principles of Method.

Analyte	Calibration Standard	Method
ALT	IFCC	Colorimetric
AST	IFCC	Colorimetric
BUN	NIST ² SRM ³ #912A	Enzymatic
CRE	NIST SRM #967	Enzymatic
GGT	IFCC	Colorimetric
GLU	NIST SRM #909	Enzymatic

Quality Control

See Section 6 (Calibration and Quality Control) of the Piccolo Xpress chemistry analyzer Operator’s Manual. Performance of the Piccolo Xpress chemistry analyzer can be verified by running controls. For a list of approved quality control materials with acceptance ranges, please contact Abaxis Technical Support. Other human serum or plasma-based controls may not be compatible. Quality control materials should be stored as per the package-insert included with the controls.

If control results are out of range, repeat one time. If still out of range, call Abaxis Technical Support. Do not report results if controls are outside their labeled limits. See Piccolo Xpress chemistry analyzer Operator’s Manual for a detailed discussion on running, recording, interpreting, and plotting control results.

Waived Laboratories (US only): Abaxis recommends control testing as follows:

- at least every 30 days
- whenever the laboratory conditions have changed significantly, e.g. Piccolo moved to a new location or changes in temperature control
- when training or retraining of personnel is indicated
- with each new lot (CLIA waived tests in waived status labs)

Non-Waived Laboratories (US only): Abaxis recommends control testing to follow federal, state, and local guidelines.

And the non-waived ones will use the below:

- Performance of the Piccolo Xpress chemistry analyzer can be verified by running controls. For a list of approved quality control materials with acceptance ranges, please contact Abaxis Technical Support. Other human serum or plasma-based controls may not be compatible. Quality control materials should be stored as per the package-insert included with the controls.
- If control results are out of range, repeat one time. If still out of range, call Abaxis Technical Support. Do not report results if controls are outside their labeled limits. See the Piccolo Xpress chemistry analyzer Operator's Manual for a detailed discussion on running, recording, interpreting, and plotting control results.

9. Results

The Piccolo Xpress chemistry analyzer automatically calculates and prints the analyte concentrations in the sample. Details of the endpoint and rate reaction calculations are found in the Piccolo Xpress chemistry analyzer Operator's Manual.

Interpretation of results is detailed in the Operator's Manual. Results are printed onto result cards or paper rolls supplied by Abaxis. The result cards or paper rolls have an adhesive backing for easy placement in the patient's files.

The reaction for each analyte occurs at 37°C (98.6°F).

10. Limitations of Procedure

General procedural limitations are discussed in the Piccolo Xpress chemistry analyzer Operator's Manual.

- The only anticoagulant **recommended for use** with the Piccolo Xpress chemistry analyzer is **lithium heparin**. Do not use sodium heparin.
- Abaxis has performed studies demonstrating that EDTA, fluoride, oxalate, and any anticoagulant containing ammonium ions will interfere with at least one chemistry contained in the Piccolo General Chemistry 6 reagent disc.
- Samples with hematocrits in excess of 62% packed red cell volume (a volume fraction of 0.62) may give inaccurate results. Samples with high hematocrits may be reported as hemolyzed. These samples may be spun down to get plasma then re-run in a new reagent disc.
- **Any result for a particular test that exceeds the assay range should be analyzed by another approved test method or sent to a referral laboratory. Do not dilute the sample and run it again on the Piccolo Xpress chemistry analyzer.**

Warning: Extensive testing of the Piccolo Xpress chemistry analyzer has shown that, in very rare instances, sample dispensed into the reagent disc may not flow smoothly into the sample chamber. Due to the uneven flow, an inadequate quantity of sample may be analyzed and several results may fall outside the reference ranges. The sample may be re-run using a new reagent disc.

Note: Operators shall report any serious incident that has occurred in relation to the device to the manufacturer.

Interference

Substances were tested as interferents with the analytes. Human serum pools were prepared. The concentration at which each potential interferent was tested was based on the testing levels in CLSI EP7-P.³⁶

Effects of Endogenous Substances

- Physiological interferents (hemolysis, icterus and lipemia) cause changes in the reported concentrations of some analytes. The sample indices are printed on the bottom of each printout to inform the operator about the levels of interferents present in each sample.
- The Piccolo Xpress chemistry analyzer suppresses any results that are affected by >10% interference from hemolysis, lipemia or icterus. "HEM", "LIP", or "ICT" respectively, is printed on the printout in place of the result.

- For maximum levels of endogenous substances contact Abaxis Technical Service.

Effects of Exogenous and Therapeutic Substances

Thirty-five exogenous and therapeutic substances were selected as potential interferents for Abaxis test methods based on recommendations by Young.³⁷ Significant interference is defined as a >10% shift in the result for a normal range specimen. Human serum pools were supplemented with a known concentration of the drugs or chemicals and then analyzed. **Please see Table 2 for a list of exogenous and therapeutic substances evaluated. Please see Table 3 for a list of analytes where interference was observed.**

Table 2: Exogenous & Therapeutic Substances Evaluated

	Highest Concentration Tested (mg/dL)
Acetaminophen	100
Acetoacetate	102
Acetylsalicylic acid	50
Ampicillin	30
Ascorbic Acid	20
Caffeine	10
Calcium Chloride	20
Cephalothin (Keflin)	400
Chloramphenicol	100
Cimetidine	16
L-Dopa	5
Dopamine	19
Epinephrine	1
Erythromycin	10
Glutathione	30
Ibuprofen	50
Isoniazide	4
α -Ketoglutarate	5
Ketoprofen	50
Methicillin	100
Methotrexate	0.5
Metyldopa	0.5
Metronidazole	5
Nafcillin	1
Nitrofurantoin	20
Oxacillin	1
Oxaloacetate	132
Phenytoin	3
Proline	4
Pyruvate	44
Rifampin	1.5
Salicylic Acid	25
Sulfasalazine	10
Sulfanilamide	50
Theophylline	20

- The following substances showed greater than 10% interference. Significant interference is defined as >10% shift in the result for a normal range specimen. Human serum pools were supplemented with known concentrations of the drugs or chemicals and then analyzed.

Table 3: The following substances showed greater than ± 10 % shift in the result for a normal range specimen

Concentration which produces > 10% Interference (mg/dL)	
Alanine Aminotransferase (ALT)	
Ascorbic acid	20
Oxaloacetate	132
Creatinine (CRE)	
Ascorbic acid	20
Dopamine	19
L-dopa	5
Epinephrine	1
Glutathione	30
Glucose (GLU)	
Oxaloacetate	132
Pyruvate	44

For additional information on potential chemical interferents, see the Bibliography.

11. Expected Values

Samples from a total of 193 adult males and females, were used to determine the reference ranges for ALT, creatinine, glucose, and BUN. Samples from a total of 186 adult males and females were used to determine the reference range for AST. Samples from a total of 131 adult males and females were used to determine the reference range for GGT. These ranges are provided as a guideline only. It is recommended that your office or institution establish normal ranges for your particular patient population.

Table 4: Piccolo Reference Intervals

Analyte	Common Units	SI Units
Alanine Aminotransferase (ALT)	10-47 U/L	10-47 U/L
Aspartate Aminotransferase (AST)	11-38 U/L	11-38 U/L
Creatinine (CRE)	0.6-1.2 mg/dL	53-106 μ mol/L
Gamma Glutamyltransferase (GGT)	5-65 U/L	5-65 U/L
Glucose (GLU)	73-118 mg/dL	4.05-6.55 mmol/L
Blood Urea Nitrogen (BUN)	7-22 mg/dL	2.5-7.9 mmol urea/L

*Data was generated using equivalence device Piccolo Blood Chemistry Analyzer.

12. Performance Characteristics

Linearity

The chemistry for each analyte is linear over the dynamic range listed below when the Piccolo Xpress chemistry analyzer is operated according to the recommended procedure (refer to the Piccolo Xpress chemistry analyzer Operator's Manual).

Table 5: Piccolo Dynamic Ranges

Analyte	Common Units	SI Units
Alanine Aminotransferase (ALT)	5-2000 U/L	5-2000 U/L
Aspartate Aminotransferase (AST)	5-2000 U/L	5-2000 U/L
Creatinine (CRE)	0.2-20 mg/dL	18-1768 µmol/L
Gamma Glutamyltransferase (GGT)	5-3000 U/L	5-3000 U/L
Glucose (GLU)	10-700 mg/dL	0.56-38.9 mmol/L
Blood Urea Nitrogen (BUN)	2-180 mg/dL	0.7-64.3 mmol/urea/L

If the analyte concentration is above the measuring range (dynamic range), but less than the system range, the printout will indicate a ">" sign at the upper limit and an asterisk after the number, e.g. ALT >2000* U/L. If lower than the dynamic range, a "<" will be printed with an asterisk, e.g. ALT <5* U/L. For values that are grossly beyond the measurement range (system range), "~~~" will be printed instead of a result. Any time "~~~" appears on a printout, collect a new sample and rerun the test. If results for the second sample are suppressed again, please call Abaxis Customer Support.

Sensitivity (Limits of Detection)

The lower limit of the reportable (dynamic) range for each analyte is: alanine aminotransferase 5 U/L; aspartate aminotransferase 5 U/L; creatinine 0.2 mg/dL (18 µmol/L); gamma glutamyltransferase 5 U/L; glucose 10 mg/dL (0.56 mmol/L); and blood urea nitrogen 2.0 mg/dL (0.7 mmol urea/L).

Precision

Precision studies were conducted using CLSI EP5-T2 guidelines.⁴² Results for within-run and total precision were determined by testing two levels of control material. Controls were run in duplicate twice each day for 20 days over a four-week period. Results of the precision studies are shown in Table 6.

Table 6: Precision (N=80)

Analyte	Within-Run	Total
Alanine Aminotransferase (U/L)		
<u>Control Level 1</u>		
Mean	21	21
SD	2.76	2.79
%CV	13.4	13.5
<u>Control Level 2</u>		
Mean	52	52
SD	2.70	3.25
%CV	5.2	6.2
Aspartate Aminotransferase (U/L)		
<u>Control Level 1</u>		
Mean	47	47
SD	0.98	1.84
%CV	2.1	3.9
<u>Control Level 2</u>		
Mean	145	145
SD	1.83	4.62
%CV	1.3	3.2

Table 6 Precision (N=80) (cont.)

Analyte	Within-Run	Total
Creatinine (µmol/L)		
<u>Control Level 1</u>		
Mean	97.24	97.24
SD	12.38	12.38
%CV	12.5	13.1
<u>Control Level 2</u>		
Mean	459.68	459.68
SD	20.33	23.87
%CV	4.4	5.2
Gamma Glutamyltransferase (U/L)		
<u>Control Level 1</u>		
Mean	25	25
SD	0.59	0.74
%CV	2.34	2.94
<u>Control Level 2</u>		
Mean	106	106
SD	1.52	2.29
%CV	1.43	2.15
Glucose (mmol/L)		
<u>Control Level 1</u>		
Mean	3.66	3.66
SD	0.04	0.06
%CV	1.1	1.6
<u>Control Level 2</u>		
Mean	15.43	15.43
SD	0.14	0.21
%CV	0.9	1.4
Blood Urea Nitrogen (mmol/L)		
<u>Control Level 1</u>		
Mean	6.78	6.78
SD	0.12	0.14
%CV	1.9	2.1
<u>Control Level 2</u>		
Mean	23.21	23.21
SD	0.38	0.42
%CV	1.6	1.8

Correlation

Heparinized whole blood and serum samples were collected from patients at two sites. The whole blood samples were analyzed at the field sites and the serum samples were analyzed by comparative methods. In two cases, the results of testing serum samples by the Piccolo were used and these are indicated appropriately in the table. In some cases, high and low supplemented samples were used to cover the dynamic range. All samples were run in singlicate on the same day. Representative correlation statistics are shown in Table 7.

Table 7: Correlation with Comparative Methods*

	Correlation Coefficient	Slope	Intercept	SEE	N	Sample Range	Comparative Method
Alanine Aminotransferase (U/L)	0.981	0.905	1.3	3.21	86	10-174	Paramax®
	0.985	0.946	-2.5	2.84	67	10-174	Technicon
Aspartate Aminotransferase (U/L)	0.93	0.87	5.3	2.76	159	13-111	Paramax
	1.0	0.97	3.0	1.9	46	13-252	DAX™
Creatinine (µmol/L)	0.993	0.926	0.0	0.15	260	35.36-1299.48	Paramax
	0.987	0.866	0.1	0.16	107	35.36-663	Beckman
Gamma Glutamyl-transferase (U/L)	1.0	0.98	-0.4	3.29	135	5-312	Paramax
	1.0*	1.60	3.1	18.57	49	27-1848	Beckman
Glucose (mmol/L)	0.987	1.009	-2.8	3.89	251	4.0-23.43	Paramax
	0.997	0.943	1.2	4.69	91	3.11-35.86	Beckman
Urea Nitrogen (mmol/L)	0.964	0.923	0.5	1.08	251	2.14 –18.56	Paramax
	0.983	0.946	0.0	0.66	92	2.14-13.57	Beckman

* One site ran only serum on the Piccolo analyzer for the gamma glutamyltransferase correlation.

**Data was generated using equivalence device Piccolo Blood Chemistry Analyzer.

Results of Untrained User Study

An “untrained user” study was conducted in which participants were given only the test instructions and asked to perform testing of 3 discs with blinded randomized samples. The samples consisted of serum pools prepared at three levels for each of the thirteen analytes: ALT, AST, creatinine, GGT, glucose, and BUN. The participants were not given any training on the use of the test. A total of approximately 60 participants were enrolled from 3 sites, representing a diverse demographic (educational, age, gender, etc) population.

Tables below present the summary of the performance for each analyte.

Alanine Aminotransferase (ALT)

	Level 1	Level 2	Level 3
N	62	62	62
Mean	45.4 U/L	98.9 U/L	184.3 U/L
%CV	3.7%	1.7%	1.5%
Observed Range	42 – 53	96 – 103	175 – 191
Percent of Results in the Range ± 15.0%*	98.4% 61/62 95%CI: 91.3% to 100%	100% 62/62 95%CI: 94.2% to 100%	100% 62/62 95%CI: 94.2% to 100%

* This percent is based on the premise that one cannot distinguish properly between normal and abnormal values when errors are greater than one-quarter of the normal range. The range of (10 U/L - 47 U/L) was considered.

Aspartate Aminotransferase (AST)

	Level 1	Level 2	Level 3
N	62	62	62
Mean	56.0 U/L	120.4 U/L	276.3 U/L
%CV	2.4%	1.1%	1.0%
Observed Range	54 – 60	117 – 124	266 – 285

Percent of Results in the Range $\pm 15.0\%$	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%
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Creatinine (CRE)

	Level 1	Level 2	Level 3
N	62	62	62
Mean	78.68 µmol/L	182,99 µmol/L	609.08 µmol/L
%CV	11.0	5.0	1.6
Observed Range	61.88 – 106,08	159.12 – 203.32	574.6 – 636.48
Percent of Results in the Range ± 15.0%	93.6 58/62 95% CI: 84.3% to 98.2%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%

Gamma Glutamyltransferase (GGT)

	Level 1	Level 2	Level 3
N	62	62	62
Mean	35.0 U/L	86.2 U/L	131.3 U/L
%CV	2.8%	1.5%	1.5%
Observed Range	33 – 38	83 – 90	123 – 135
Percent of Results in the Range ± 15.0%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%

Glucose (GLU)

	Level 1	Level 2	Level 3
N	62	62	62
Mean	5.28 mmol/L	7.23 mmol/L	21.31 mmol/L
%CV	1.1%	1.0%	0.8%
Observed Range	5.16 – 5.44	6.94 – 7.38	19.48 – 20.71
Percent of Results in the Range ± 10.4%**	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%

** The range of (65 mg/dL - 99 mg/dL) was considered.

Blood Urea Nitrogen (BUN)

	Level 1	Level 2	Level 3
N	62	62	62
Mean	5.61 mmol/L	14.64 mmol/L	25.78 mmol/L
%CV	2.3	2.5	1.8
Observed Range	5.0 – 5.71	13.21 – 15.35	24.28 – 26.78
Percent of Results in the Range ± 15.0%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%	100% 62/62 95% CI: 94.2% to 100%

13. Symbols



Use By



Catalog Number



Batch Code



In Vitro Diagnostic
Medical Device



Consult Instructions
for Use



Manufacturer



Do Not Reuse



X Number of Test
Devices In Kit



Manufacturing
Sequence



Serial Number



Authorized
Representative
in the European
Community



Temperature
Limitation



PN:
Part Number

Caution



denotes conformity to specified
European directives

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